## **Evaluation of MycoSEQ Plus Method for Rapid Mycoplasma Contamination Detection**

**Senior Associate Scientist Daniel Berry** 

ARD MST (Analytical Research and Development Microbiology Strategy and Testing), Pfizer Inc. Andover MA

Acknowledgments: Nasrin Salehi, Michelle Norton, Nate Stewart

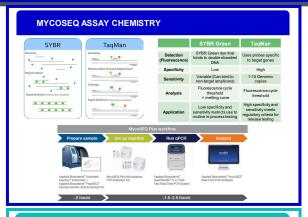
## MYCOPLASMA CONTAMINATION DETECTION

Testing for mycoplasma contamination in cell banks and bioreactor cell cultures is a regulatory requirement for production of biological products. The compendial tests for mycoplasma detection require 14-28 days incubation, imposing a limitation on batch release timelines and rapid containment in the event of a contamination. The alternative rapid PCR-based mycoplasma detection kit, MycoSEQ Plus, provides results within a few hours and meets regulatory guidelines regarding sensitivity (10 CFU/mL or the genomic equivalent of 10 GC/mL) and specificity as outlined in the European Pharmacopoeia (E.P. 2.6.7, 2007), US Pharmacopoeia (US63), and Japanese Pharmacopoeia.

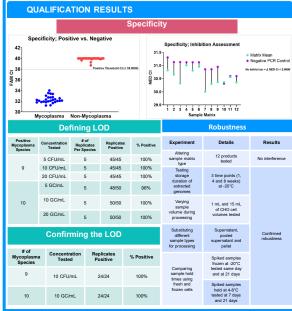
## Wycopiasma Tosting Points and Approaches Upstream Proces Downsteam Process 15 no. 25 no. 10 points and 25 no. 1

		Culture Methods	PCR	Key Differences	
1	Test Method	Agar, Broth and Cell Culture	Real-Time PCR	PCR does not require cultivation, therefore limited impact of biocidal or bacteriostatic compounds.	
	Specificity	High	High	PCR uses primers specific to mycoplasma and eliminating the need for multiple culture methods.	
	Sensitivity	10-100 CFU/mL	10 CFU/mL	Equivalent sensitivity.	
R	Sample equirements	15 mL	15 mL	Equivalent sample requirement.	
	Timeline	~28 days	Hours	Significantly reduced timeline.	
Spe	ecies Detected	Viable Mycoplasma	> 200 species	PCR can detect Mycoplasma contamination regardless of viability.	





Para	ameter	USP <1223>						
Spe	cificity	Ability to detect organism with no interference from sample						
Limit of Detection (LOD)		Lowest number of organisms that can be detected						
Robi	ustness	A measure of method's capacity to remain unaffected by small but deliberate variations in method						
Rugg	jedness	Challenge with different analysts, reagent lots, instruments, etc.						
	LOD							
Mycoplasma Strains	Non-Mycoplasma Strains	Cell Culture Matrix (Assess Inhibition)	Inactivated Mycoplasma Stock	Genom		# of Replicates		
Required	Closely related bacteria to Mycoplasma required by regulatory Clostridium sporogenes Lactobaciflus areas Streptococcus salivatium acetobutylicum acetobutylicum Bacillus subilis	12 different products from different modalities	5 CFU/mL	5 GC/mL		5		
Mycoplasma strains by regulatory in CFU			10 CFU/mL	10 GC/mL		5 and 24		
and GC titer			20 CFU/mL	20 GC/mL		5		
Acholeplasma laidlawii Mycoplasma arginini			Robustness					
Mycoplasma fermentans Mycoplasma gallisepticum dycoplasma hominis fycoplasma hyorhinis Mycoplasma orale			Variations		Testing			
			Sample Matrix Interface		12 Products			
			Hold Time Storage of extracted genome		3 Time Points			
Mycoplasma pneumoniae	Pseudomonas aeruginosa		Sample Volume		2 Volumes			
Mycoplasma	Micrococcus luteus Stenotrophomonas		Alternate Sample Type		3 Types			
salivarium Mycoplasma synoviae	maltophilia Foterobacter cloacae		Sample Hold Time		2 Storage Conditions and 2 Time Points each			



## CONCLUSION

The assay can be used as a platform assay in CHO products. The MycoSEQ plus assay generates results in a few hours with a LOD of 10CFU or GC/mL, meeting regulatory criteria. Our testing demonstrated no interactions with other bacteria or sample matrices. We have concluded that this assay is suitable as a platform method for mycoplasma detection for in-process and release testing of bioproduction samples.